



Tropentag 2020,
September 9-11, 2020

virtual conference

Conference on International Research on Food Security, Natural Resource
Management and Rural Development
organised by ATSAF e.V., Germany

Gaseous N and C losses during sun-drying of goat manure: *Effects of drying conditions and feed additives*

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Abstract

Animal manure is a key resource in farming systems of arid and semi-arid regions. Its quality is often low due to inappropriate storage. Thus, this study aimed to assess the effects of drying and storage conditions on nutrient losses during typical sun-drying of manure in the Sultanate of Oman, and how these losses can possibly be mitigated by changes in manure properties. Charcoal and tannins are known to stabilize organic matter and increase nitrogen (N) retention in soils and compost. Therefore, they were used as feed additives to stabilize carbon (C) and N in manure.

NH₃-N, N₂O-N, and CO₂-C losses from sun-drying goat manure were measured in three experiments during the cropping season, differing in temperatures and resulting drying rates. Manure was obtained from goats fed a diet of 50% hay, 47% maize and 3% soybean with or without the addition of either 2.6% activated charcoal (AC) or 3.4% Quebracho tannin extract (QT).

The cumulative N and C losses reached up to 1.4% and 2.2% of the initial N and C contents, respectively. During the drying process, control manure lost 138 to 295 mg N kg⁻¹, AC lost 112 to 207 and QT 107 to 208 mg kg⁻¹ via NH₃ volatilization and N₂O emissions. Carbon losses during drying ranged from 0.1 to 10.2, 0 to 10.5, and 0.3 to 5.4 g kg⁻¹ for Control, AC and QT manure, respectively. Slow drying (up to 84 h) favored CO₂ emissions and reduced NH₃ volatilization due to higher microbial activity and an immobilization of mineral N in manure. In comparison, two times more NH₃-N was lost during quick drying of manure (4 h) than during slow drying, even after manure reached constant weight. In contrast to dietary charcoal, dietary tannins reduced N and C losses by up to 60% during manure drying and storage, potentially increasing its quality as organic fertilizer and improving nutrient recycling.

Key words: Ammonia, nitrous oxide, carbon dioxide, goat manure, activated charcoal, tannin, Oman

Introduction

In many parts of the tropics and subtropics crop production still strongly relies on animal manure as a nutrient source particularly in low input systems (*Rufino et al., 2007; Buerkert et al., 2010*). In traditional oasis agriculture of Oman, grazing animals leaving their droppings in stables overnight contribute to accumulating nutrients from huge grazing areas within the crop fields of small villages (*Schlecht et al., 2011*). However, since these droppings are often dried in the sun, collected and stored in plastic bags or left in uncovered heaps until used as manure, considerable amounts of carbon (C) and nitrogen (N) are lost (*Buerkert et al., 2010*). These losses are not only relevant for nutrient cycling within farming systems, but may also lead to environmental hazards such as soil acidification and greenhouse gas emissions. Gaseous emissions from manure depend on the moisture content of stored manure, ambient temperatures and manure properties, such as the C/N ratio (*Liang et al., 2003; Buerkert et al., 2010*).

The adaptation of animal diets offers a way to control emissions from manure as the diet directly affects the amount and composition of manure (*Petersen et al., 2007*). For instance, reducing dietary N intake decreases NH₃ and N₂O emissions from stored dairy cow manure (*Külling et al., 2001*) and from soil incubated with manure (*Lee et al., 2013*) by reducing urinary N excretion and increasing C/N ratio of feces. Therefore, feed additives affecting C and N digestibility of diets such as charcoal or tannins are likely to affect gaseous C and N losses from manure.

Charcoal is used in animal nutrition to improve animal health, reduce greenhouse gas emissions from ruminants and deodorize manure (*Poage et al., 2000; Watarai et al., 2008*). As a stable C source, the addition of 3% activated charcoal to goats' diets can increase manure C by 12% and the C/N ratio by 26% (*Ingold et al., 2015; Al-Kindi et al., 2016*). Tannins naturally occur in many browse species (*Muir, 2011*) and have a high affinity to bind and precipitate proteins (*Spencer et al., 1988*). This property is responsible for diverse effects in animal nutrition and health, such as reduced methane emission and bloat, increased milk production and growth rates, but also reduced fodder digestibility, toxicity and an altered nitrogen use efficiency and excretion (*Aerts et al., 1999; Puchala et al., 2005; Makkar, 2003; Al-Kindi et al., 2016*). How changes of manure composition due to feed additives such as activated charcoal or tannins affect C and N losses during manure drying requires further research. To fill this research gap three experiments were conducted that tested the effect of activated charcoal (AC) and quebracho tannins (QT) as feed additives on gaseous C and N losses from manure under differing climatic conditions in Northern Oman. We hypothesized that quick drying of manure in the sun reduces C and N losses compared to slow drying, and that AC and QT feed additives stabilize C and N in manure, reducing gaseous C and N losses during drying and storage.

Material and Methods

The experiments were conducted on a private farm near Sohar (24°24' N, 56°79' E) in the Al-Batinah Plain of the Sultanate of Oman. Experiment 1 was conducted under hot temperatures with high humidity (24.4 to 45.3°C, and 36 to 84% relative humidity). Experiment 2 and 3 were conducted at lower temperatures (18.9 to 39.4°C and 12.0 to 28.9°C, respectively) and differing humidity levels (9 to 72% and 44 to 82% relative humidity, respectively). Manure was collected from male Jebel Akhdar goats fed a basal diet of 50% Rhodes grass hay (*Chloris gayana* Kunth), 46.5% crushed maize (*Zea mays* L.) and 3.5% soybean meal (*Glycine max* (L.) Merr.; Ingold et al., 2015). The 60 goats were divided into three groups receiving only the basal diet (control [Co]), or additions of 2.5% of a steam-activated charcoal on coconut shell basis (AquaSorb® CP1, Jacobi Carbons Service (Europe) GmbH, Premnitz, Germany) or 3.6% of a water soluble quebracho tannin extract (*Schinopsis balansae* Engl.; Silva Team S.p.a., San Michele Mondovì CN, Italy), respectively. The collected feces were pooled per treatment and kept in plastic bags for a maximum of two hours before sun drying. Fresh feces were spread in PVC containers of 30 cm diameter in three replicates per treatment at 1.4, 10.8 and 8.8 kg m⁻² in experiment 1, 2 and 3, respectively. The differing manure quantities resulted in fast drying of manure in experiment 1 (< 4 h) and slow drying (> 80 h) in experiment 2 and 3. Fresh manure contained 56, 52 and 62% moisture, 2.17, 1.88 and 2.26% N, 45.5, 49.3 and 46.2% C and had a pH of 6.0, 6.4 and 6.4 for

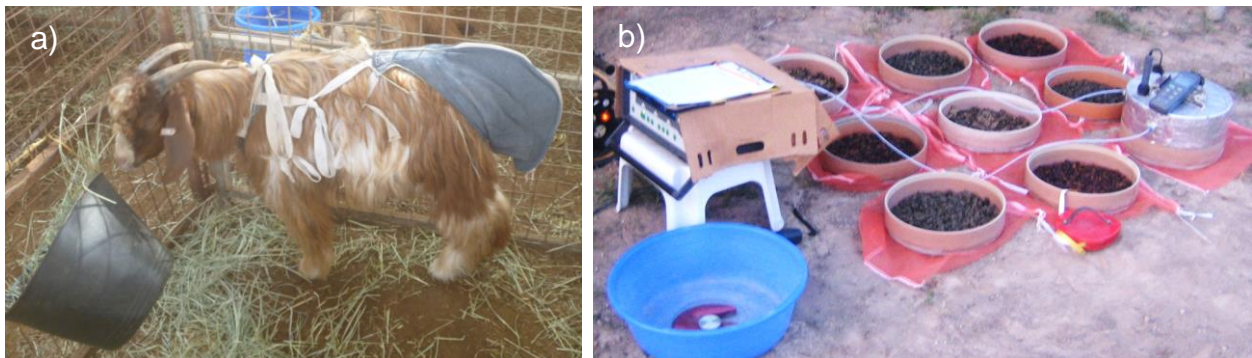


Figure 1: Goat with fecal collection bag in individual crate (a) and in-field gas emission measurement from drying manure using a closed chamber connected to a photo-acoustic multi-gas analyzer in Sohar, Sultanate of Oman (b).

Co, AC and QT, respectively.

NH₃-N volatilization and N₂O-N and CO₂-C emissions during the drying process were measured with a dynamic closed-chamber system using a photo-acoustic infrared multi-gas monitor (INNOVA 1312-5, LumaSense Technologies A/S, Ballerup, Denmark; Predotova et al., 2010). The PVC containers were covered with a Teflon® coated PVC cuvette for four minutes, which was attached to the multi-gas monitor by two 1.5 m long Teflon® tubes for in- and out-coming gas. Gas fluxes were calculated using linear regression in the R package “gasfluxes” (Fuss, 2017). Linear regressions with R² < 0.6 were set to zero. Cumulative emission during the drying process (until manure dry matter reached ≥90%) were calculated by linear interpolation of consecutive

data points multiplied by the time interval. Gas emission rates were statistically analyzed by generalized linear regressions with individual PVC containers as repeatedly measured subjects and treatment and drying condition ($<$ and $\geq 90\%$ DM) as fixed factors. In case of non-normal distribution of residuals, a non-parametric Kruskal-Wallis-Test was conducted. Cumulative N and C losses were analysed by ANOVA followed by Tukey-HSD test. All statistical analyses were conducted with IBM SPSS Statistics, Version 24 (IBM Deutschland GmbH, Nürnberg, Germany).

Results and Discussion

NH_3 -N volatilization and N_2O -N and CO_2 -C emission were highest directly after spreading manure in PVC containers and declined thereafter with decreasing moisture content of manure until constantly low levels (data not shown). The gaseous N and C losses during the drying process in the sun amounted to 0.5 to 1.4% of initial N and 0.01 to 2.2% of initial C content of manure, respectively. They were thus in a similar range as N losses from unroofed manure storage in Niger (Predotova et al., 2010). However, cumulative C and N losses from manure conservation and storage are reported to be much higher with up to 50% of initial C and N being lost (Chadwick, 2005; Predotova et al., 2010; Tiftonell et al., 2010). In contrast to these studies, manure spread on the ground for sun-drying under the hot and sunny conditions of Oman resulted in very quick drying of manure. When estimating possible C losses during manure storage for 50 days based on emission rates of dried manure, losses amounted to 0.01 to 3.2% of initial manure C. While cumulative N losses over a 50-day storage period in experiments 2 and 3 amounted to 1 to 3.4% of initial manure N; in experiment 1 N losses were estimated to reach 35 to 62%. Depending on the ammonium concentration of manure - the main source of NH_3 volatilization - N losses during longer periods of manure storage can be considerable.

Effect of drying conditions

In experiment 1 average NH_3 volatilization and N_2O emission rates from manure were up to 39-times higher than in experiments 2 and 3, whereas average CO_2 emission rates were up to 48-times lower (Figure 2). The major difference between experiments 1, 2 and 3 was the different quantity of manure per m^2 resulting in slower drying of manure in experiments 2 and 3 (> 80 h until $\text{DM} \geq 90\%$ compared with 4 h in experiment 1). Correspondingly, manure storage under rainy conditions reduces gaseous N losses and increases gaseous C losses compared to dry conditions (Chadwick, 2005, Predotova et al., 2010; Buerkert et al., 2010). CO_2 evolution is a sign of microbial respiration and indicates the activity of microbial biomass in manure, which is mainly affected by moisture content and temperature (Liang et al., 2003; Buerkert et al., 2010). Multiple linear regression analysis explained 78% of the variance in CO_2 emissions from drying manure and demonstrated the impact of manure moisture content on CO_2 emissions, whereas temperature was less influential (Table 1). In contrast, the linear model of the physical process of NH_3 volatilization explained 51% of the variance and was driven by manure moisture and air

temperature; it negatively correlated with CO₂ emission rates. The negative impact of CO₂ emission rates on NH₃ volatilization, included in the model as an indicator of microbial activity, indicated an immobilization of ammonium N in manure due to synthesis of microbial biomass N.

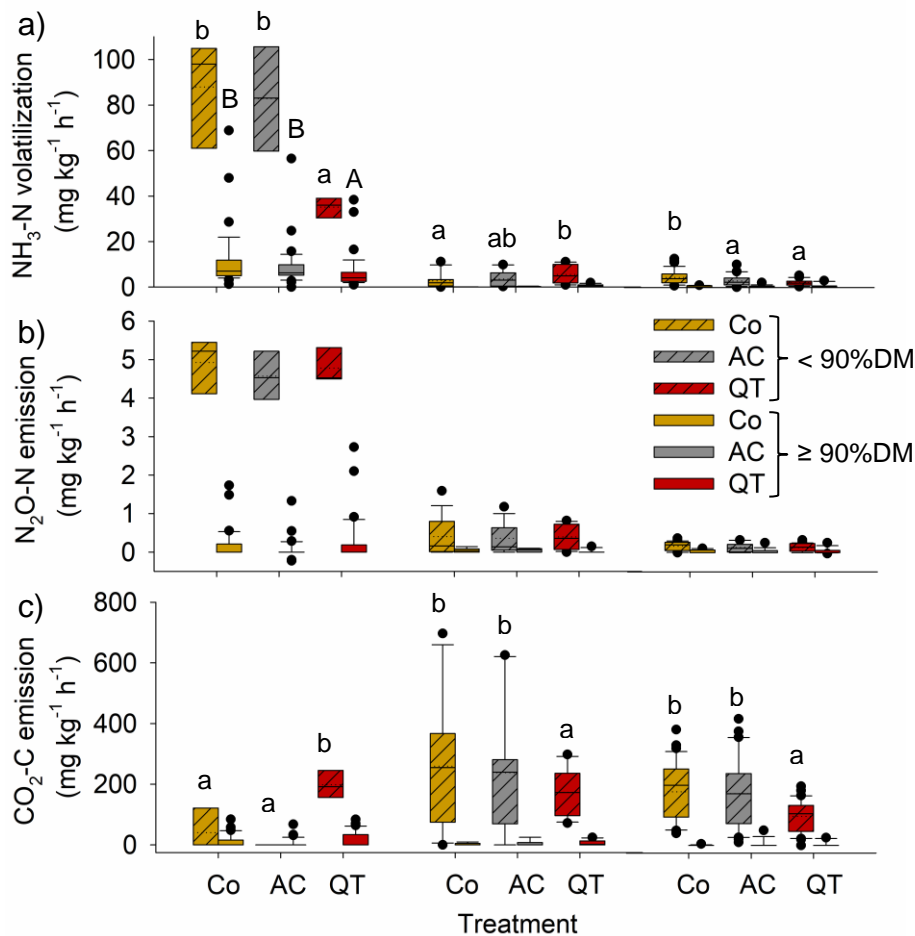


Figure 2: Boxplots of NH₃-N (a), N₂O-N (b), and CO₂-C (c) emission rates from drying manure (< 90% DM) and after reaching constant weight (≥ 90% DM) measured in three experiments in Sohar, Oman. Small letters indicate significant effects between manure treatments < 90% DM and capital letters significant differences between manure treatments ≥ 90% DM.

Treatment effects

Charcoal-enriched manure (AC) and tannin-enriched manure (QT) may be considered a by-product of the usage of charcoal or tannin-containing fodder in order to improve animal nutrition and health (Aerts et al., 1999; Poage et al., 2000; Makkar, 2003; Puchala et al., 2005; Watarai et al., 2008). Feeding charcoal as a C-rich substrate and tannins, known to protect proteins from digestive breakdown, was expected to stabilize C and N compounds in manure by lowering the gaseous C and N losses from manure. The treatments significantly affected the daily CO₂ emission rate and NH₃ volatilization during drying (P = 0.003 and 0.041, respectively), whereas N₂O emissions were generally low and unaffected by treatments (Figure 2).

Table 1: Linear model of predictors (time of drying, manure moisture and temperature) of $\text{NH}_3\text{-N}$, $\text{N}_2\text{O-N}$ and $\text{CO}_2\text{-C}$ emission rates ($\text{mg h}^{-1} \text{kg}^{-1}$) during manure drying of the three experiments conducted in Sohar, Sultanate of Oman, with non-standardised and standardised beta coefficients, standard errors (SE), significance levels (p) and coefficients of determination (R^2).

		Non-standardised		Standardised	p	Overall	
		b	SE	β		p	R^2
CO_2	Constant	-119.8	20.99		<0.001	<0.001	0.78
	Moisture	4.8	0.23	0.77	<0.001		
	Temperature	3.2	0.64	0.18	<0.001		
N_2O	Constant	-1.55	0.19		<0.001	<0.001	0.34
	Moisture	0.02	0.00	0.44	<0.001		
	Temperature	0.05	0.01	0.41	<0.001		
NH_3	Constant	-28.08	3.24		<0.001	<0.001	0.51
	Time	-0.05	0.02	-0.16	0.003		
	Moisture	0.57	0.05	0.82	<0.001		
	Temperature	1.03	0.08	0.53	<0.001		
	CO_2 emission rate	-0.09	0.01	-0.82	<0.001		

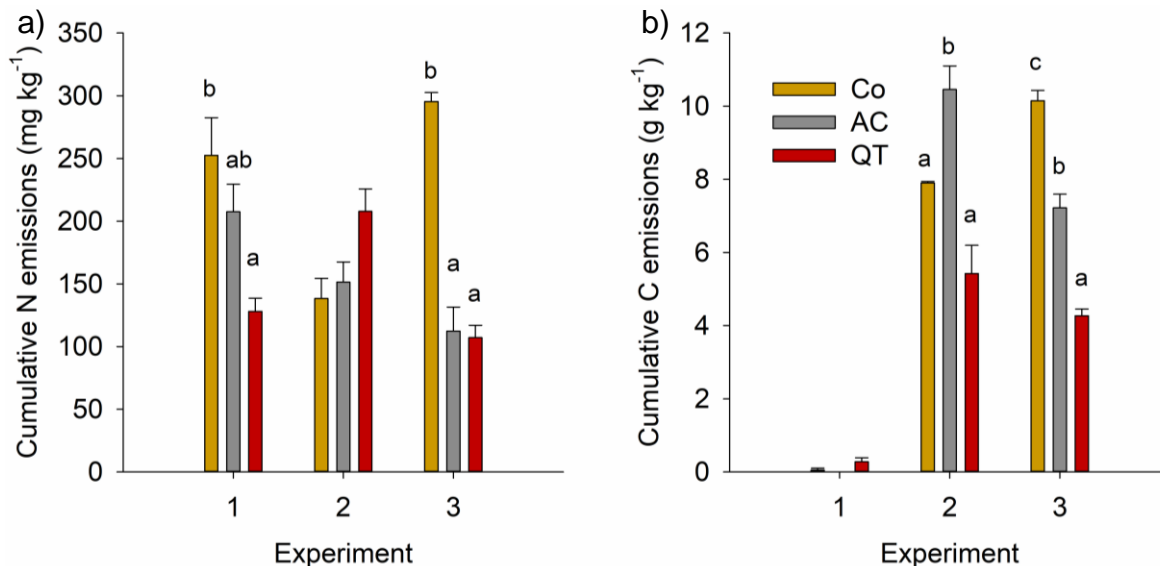


Figure 3 Cumulative N (a) and C emissions (b) during drying of manure in three experiments conducted in Sohar, Sultanate of Oman. Letters indicate significant treatment effects within the three experiments.

Charcoal increased fecal C concentration and reduced fecal N concentration resulting in higher fecal C/N ratios compared with unamended control manure (Ingold et al., 2015; Al-Kindi et al., 2016). Surprisingly, during drying of manure and storage thereafter, no clear effects of dietary AC on the measured gas emission rates were observed. However, cumulative N losses were reduced in two experiments by 18 to 62% (Figure 3), which showed the N conservation potential of AC. Higher C/N ratios and the high sorption capacity of charcoal can lead to an immobilization of N, thus reducing gaseous N losses in form of N_2O and NH_3 (Cayuela et al., 2013; Maurer et al., 2017). Cumulative C losses were inconsistently affected by AC in the three experiments, which was also reported in a recent review on manure amendment of biochar (Maurer et al., 2017).

Quebracho tannins used as feed additives slightly increased C and N concentrations in feces, resulting in similar C/N ratios of about 20:1 (Ingold et al., 2015; Al-Kindi et al., 2016). During drying

and subsequent storage, NH₃ volatilization rates were significantly reduced by 43 to 54% with QT in experiments 1 and 3 compared to Co (Figure 2). CO₂ emissions were differently affected in the three experiments. When manure dried quickly, like in experiment 1, QT emitted on average 4- and 20-fold more CO₂ than AC and Co manure, respectively, although emissions were generally low.

Manure moisture content was found to be the main driver of CO₂ emissions from drying manure, which was with 62% by 11% and 19% higher than in AC and Co, respectively. The higher initial moisture content in manure provided more favorable conditions for microbial activity, measured as CO₂ emission. In experiments 2 and 3, manure dried much slower than in experiment 1, allowing for high microbial activity over several days. Under these conditions, QT reduced CO₂ emissions by 40 to 60% (Figure 3). Tannins are known for their antimicrobial properties (Scalbert, 1991) and were found to reduce microbial respiration in soils and compost (Jordan et al., 2015), although this inhibitory effect was not always observed (Hao et al., 2000). The activity depression can be caused by direct toxic effects on microorganisms or enzyme inhibition due to binding of tannins to bacterial cell walls and enzymes (Haettenschwiler and Vitousek, 2000). However, after having passed through the animal's gastrointestinal tract, it is likely that tannins are bound to feces components and may therefore not actively inhibit microorganisms (Degen et al., 1995) but rather reduce the availability of easily degradable organic substrates by complexation or by a shift of N and C excretion towards more stable forms (Al-Kindi et al., 2016).

Conclusions

The results show that N and C losses from sun-drying of feces are with 0.5 to 1.4% N and 0.01 to 2.2% C of initial N and C contents, respectively, relatively low compared with composting and storage in heaps. N and C losses during subsequent storage of properly dried manure were very low, except for NH₃ volatilization losses of quickly dried manure (experiment 1). The considerable NH₃ volatilization rates of quickly drying manure point to the risk of a high loss of ammonium N from manure during storage. Slow drying, which allows microbial biomass to grow for a short time, may preserve mineral N in form of microbial biomass N within manure. While charcoal-enriched manure showed inconsistent effects on gaseous N and C losses, tannins reduced N and C losses by up to 60% compared with control manure and may be a useful tool in conserving N and C during manure drying and storage and thus increasing the efficiency of nutrient cycling within the farming system.

Acknowledgements

We thank Royal Court Affairs (Royal Gardens and Farms), Sultanate of Oman, for its support and the German Research Foundation (DFG) for funding this research within the Research Training Group 1397 "Regulation of Soil Organic Matter and Nutrient Turnover in Organic Agriculture" at Universität Kassel, Germany.

References

- Aerts, R. J., Barry, T. N., McNabb, W. C. (1999): Polyphenols and agriculture: beneficial effects of proanthocyanidins in forages. *Agr. Ecosys. Environ.* 75, 1-12.
- Al-Kindi, A., Dickhoefer, U., Schlecht, E., Sundrum, A., Schiborra, A. (2016): Effects of quebracho tannin extract (*Schinopsis balansae* Engl.) and activated charcoal on nitrogen balance, rumen microbial protein synthesis and faecal composition of growing Boer goats. *Arch. Anim. Nutr.* 70, 307-321.
- Buerkert, A., Jahn, H., Golombek, S. D., Al Rawahi, M. N., Gebauer, J. (2010): Carbon and nitrogen emissions from stored manure and cropped fields in irrigated mountain oases of Oman. *J. Agr. Rural Develop. Trop. Subtrop.* 111, 55-63.
- Cayuuela, M. L., Sánchez-Monedero, M. A., Roig, A., Hanley, K., Enders, A., Lehmann, J. (2013): Biochar and denitrification in soils: when, how much and why does biochar reduce N₂O emissions? *Sci. Rep.* 3, 1732.
- Chadwick, D. R. (2005): Emissions of ammonia, nitrous oxide and methane from cattle manure heaps: effect of compaction and covering. *Atmos. Environ.* 39, 787-799.
- Degen, A. A., Becker, K., Makkar, H. P. S., Borowy, N. (1995): *Acacia saligna* as a fodder tree for desert livestock and the interaction of its tannins with fibre fractions. *J. Sci. Food Agr.* 68, 65-71.
- Fuss, R. (2017): Greenhouse gas flux calculation from chamber measurements. Version 0.2-1. <https://cran.r-project.org/web/packages/gasfluxes/>
- Haettenschwiler, S., Vitousek, P. M. (2000): The role of polyphenols in terrestrial ecosystem nutrient cycling. *Trends Ecol. Evol.* 15, 238-243.
- Hao, X., Chang, C., Larney, F. J., Travis, G. R. (2000): Greenhouse gas emissions during cattle feedlot manure composting. *J. Environ. Qual.* 30, 376-386.
- Ingold, M., Al-Kindi, A., Jordan, G., Dietz, H., Schlecht, E., Buerkert, A. (2015): Effects of activated charcoal or quebracho tannins added to feed or as soil conditioner on manure quality. *Org. Agric.* 5, 245-261.
- Jordan, G., Predotova, M., Ingold, M., Goenster, S., Dietz, H., Joergensen, R. G., Buerkert, A. (2015): Effects of activated charcoal and tannin added to compost and to soil on carbon dioxide and ammonia volatilisation. *J. Plant Nutr. Soil Sci.* 178, 218-228.
- Külling, E. R., Menzi, H., Kröber, T. F., Neftel, A. (2001): Emissions of ammonia, nitrous oxide and methane from different types of dairy manure during storage as affected by dietary protein content. *J. Agric. Sci.* 137, 235-250.
- Lee, C., Feyereisen, G. W., Hristov, A. N., Dell, C. J., Kaye, J., Beegle, D. (2013): Effects of dietary protein concentration on ammonia volatilisation, nitrate leaching, and plant nitrogen uptake from dairy manure applied to lysimeters. *J. Environ. Qual.* 43, 398-408.
- Liang, C., Das, K. C., McCledon, R. W. (2003): The influence of temperature and moisture contents regimes on the aerobic microbial activity of a biosolids composting blend. *Bioresource Technol.* 86, 131-137.
- Makkar, H. P. S. (2003): Effects and fate of tannins in ruminant animals, adaptation to tannins, and strategies to overcome detrimental effects of feeding tannin-rich feeds. *Small Ruminant Res.* 49, 241-256.
- Mauer, D. L., Kozziel, J. A., Kalus, K., Anderson, D. S., Opalinski, S. (2017): Pilot-scale testing of non-activated biochar for swine manure treatment and mitigation of ammonia, hydrogen sulfide, odorous volatile organic compounds (VOCs), and greenhouse gas emissions. *Sustainability* 9, 929. doi:10.3390/su9060929.
- Muir, J. P. (2011): The multi-faceted role of condensed tannins in the goat ecosystem. *Small Ruminant Res.* 98, 115-120.
- Petersen, S. O., Sommer, S. G., Béline, F., Burton, C., Dach, J., Dourmad, J. Y., Leip, A., Misselbrook, T., Nicholson, F., Poulsen, H. D., Provolo, G., Sørensen, P., Vinnerås, B., Weiske, A., Bernal, M. P., Böhm, R., Juhász, C., Mihelic, R. (2007): Recycling of livestock manure in a whole-farm perspective. *Livest. Sci.* 112, 180-191.
- Poage, G. W., Scott, C. B., Bisson, M. G., Hartmann, F. S. (2000): Activated charcoal attenuates bitterweed toxicosis in sheep. *J. Range Manage.* 53, 73-78.
- Predotova, M., Schlecht, E., Buerkert, A. (2010): Nitrogen and carbon losses from dung storage in urban gardens of Niamey, Niger. *Nutr. Cycl. Agroecosys.* 87, 103-114.
- Puchala, R., Min, B. R., Goetsch, A. L., Sahl, T. (2005): The effect of a condensed tannin-containing forage on methane emission by goats. *J. Anim. Sci.* 83, 182-186.
- Rufino, M. C., Tiftonell, P., van Wijk, M. T., Castellanos-Navarrete, A., Delve, R. M., de Ridder, N., Gillter, K. E. (2007): Manure as a key resource within smallholder farming systems: Analysing farm-scale nutrient cycling efficiencies with the NUANCES framework. *Livest. Sci.* 112, 273-287.
- Scalbert, A. (1991): Antimicrobial properties of tannins. *Phytochemistry* 30, 3875-3883.
- Schlecht, E., Dickhoefer, U., Predotova, M., Buerkert, A. (2011): The importance of semi-arid natural mountain pastures for feed intake and recycling of nutrients by traditionally managed goats on the Arabian Peninsula. *J. Arid Environ.* 75, 1136-1146.
- Spencer, C. M., Cai, Y., Martin, R., Gaffney, S. H., Goulding, P. N., Magnolato, D., Lilley, T. H., Haslam, E. (1988): Polyphenol complexation – some thoughts and observations. *Phytochemistry* 27, 2397-2409.
- Tiftonell, P., Rufino, M. C., Janssen, B. H., Giller, K. E. (2010): Carbon and nutrient losses during manure storage under traditional and improved practices in smallholder crop-livestock systems – evidence from Kenya. *Plant Soil* 328, 253-269.
- Watarai, S., Tana, Koiwa, M. (2008): Feeding activated charcoal from bark containing wood vinegar liquid (nekka-rich) is effective as treatment for cryptosporidiosis in calves. *J. Dairy Sci.* 91, 1458-1463.